



## The 65th ASH Annual Meeting Abstracts

## POSTER ABSTRACTS

## 703.CELLULAR IMMUNOTHERAPIES: BASIC AND TRANSLATIONAL

**Phenotypic, Transcriptomic and Proteomic Characteristics of CAR T-Cell Dysfunction Are Associated with Inferior CAR T-Cell Expansion and Treatment Failure in r/r B-NHL**

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**Introduction:** Despite the curative potential of CAR T-cells, a significant number of patients fail to respond or relapse early. Recently, we and others showed that early expansion failure, and particularly CAR T-cell levels at day 7, are strong predictors for treatment resistance (Blumenberg et al, Blood Supplement 2022; Locke et al, Blood Adv 2020). Here we dissected early characteristics of CAR T-cell dysfunction based on immune checkpoint expression, transcriptomic signatures and cytokine profiles and investigated their link to CAR T-cell expansion failure and patient outcome of CD19-targeted CAR T-cell therapy.

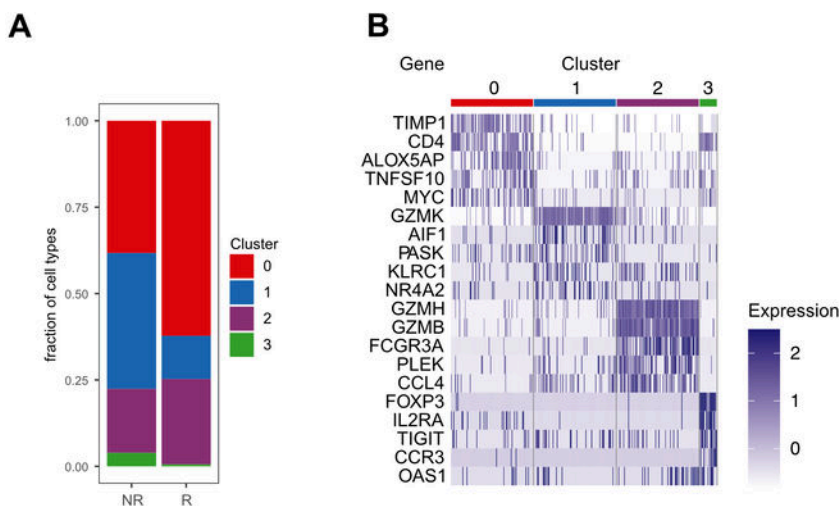
**Methods:** Patients with r/r B-NHL who underwent treatment with axi-cel or tisa-cel between January 2019 and November 2021 in the third- or later-line setting at the LMU in Munich were included (n=55). EDTA-anticoagulated peripheral blood and serum was collected. CD19 CAR and immune checkpoint expression (IC; PD-1, TIM-3, LAG-3 and CD224) was assessed by multiparameter flow cytometry on day 14 after CAR T-cell infusion. A next-generation targeted single-cell proteogenomics approach (BD Rhapsody™) to analyze the transcriptome of distinct CAR T-cell subpopulations on day 7 and day 28 after CAR T-cell infusion. A total of 92 human immuno-oncology related proteins were simultaneously measured using the Olink® Immuno-Oncology panel on day 14 after infusion. Signatures of CAR T-cell dysfunction have been compared between patients with low (loEx) or high (hiEx;  $< vs \geq 19$  CAR T-cells/ $\mu$ l at day 7 post infusion) CAR T-cell expansion and were put into context of clinical outcome measures.

**Results:** CAR T-cell levels at day 7 were inversely correlated to co-expression of PD-1 ( $r = -0.4923$ ,  $p = 0.0011$ ,  $n = 41$ ), TIM-3 ( $r = -0.3634$ ,  $p = 0.0195$ ,  $n = 41$ ) and LAG-3 ( $r = -0.5412$ ,  $p = 0.0003$ ,  $n = 41$ ) on CAR T-cells at day 14, suggestive of an association of early expansion failure and dysfunction of CAR T-cells. Indeed, loEx ( $n=26$ ) showed higher frequencies of CAR T-cells coexpressing several IC, including LAG-3<sup>+</sup> ( $p=0.0036$ ), LAG-3<sup>+</sup>PD-1<sup>+</sup> ( $p = 0.0048$ ), or LAG-3<sup>+</sup>TIM-3<sup>+</sup> ( $p = 0.0065$ ) compared to hiEx ( $n=15$ ) on day 14 after transfusion. Conversely, we detected higher frequencies of non-dysfunctional CAR T-cell phenotypes negative for several IC, such as CD244<sup>+</sup>PD-1<sup>-</sup>TIM-3<sup>-</sup> ( $p = 0.0006$ ), CD244<sup>+</sup>LAG-3<sup>-</sup>PD-1<sup>-</sup>TIM-3<sup>-</sup> ( $p = 0.001$ ) and CD244<sup>+</sup>LAG-3<sup>+</sup>PD-1<sup>-</sup>TIM-3<sup>-</sup> ( $p = 0.011$ ) in hiEx. Next, we applied uniform manifold approximation and projection (UMAP) data to examine the surface expression of IC on CAR T-cells at day 14 post-infusion according to treatment response. Unsupervised clustering revealed a higher abundance of CAR T-cell clusters positive for several IC (CD244<sup>+</sup>LAG-3<sup>-</sup>PD-1<sup>+</sup>TIM-3<sup>+</sup>) in NR ( $n = 16$ ) compared to R ( $n = 23$ ). By contrast, CAR T-cell clusters containing CAR T-cells expressing no IC were more abundant in R compared to NR. In addition, we found significantly increased levels of a highly dysfunctional CAR T-cell phenotype positive for all four IC in NR compared to R at day 14 post infusion ( $p = 0.0178$ ,  $n = 46$ ). When specifically comparing the transcriptome of CAR T-cells between NR ( $n = 6$ ) and R ( $n = 6$ ), immune inhibitory and exhaustion-related genes were

upregulated in NR, such as GZMK, KLRC1 and NR4A2 (figure 1, cluster 1). Conversely, CAR T-cells in R overexpressed genes related to T-cell effector function and cell differentiation, such as TIMP-1, TNFSF10 and MYC (figure 1, cluster 0). Consistent with these findings, a multiplexed proteomics assay for immunomodulatory serum factors in day 14 samples revealed an upregulation of proteins linked to an immune inhibitory and inflamed milieu in patients with highly dysfunctional CAR T-cells (e.g., sPD-L1 and sLAG-3,  $p = 0.0235$  and  $p = 0.0264$ , respectively,  $n = 46$ ) as well as in non-responding patients (e.g., sPD-L1 and IL-6,  $p = 0.0036$  and  $p = 0.0053$ , respectively,  $n = 55$ ).

**Conclusion:** We were able to characterize a phenotypic, transcriptomic and proteomic signature of CAR T-cell dysfunction in post-infusion samples of both patients experiencing CAR T-cell expansion failure as well as lack of radiographic response. Our data show that treatment failure might be predicted not only by CAR T-cell kinetics but also by early assessment of the CAR T-cell phenotype, transcriptome and secretome, and thus enables identification of patients in need of salvage treatment.

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**Figure 1. Transcriptomic correlates of CAR T-cell dysfunction.** (A) Bar plot indicating fraction of cell types in non-responding (468 cells) and responding (368 cells) patients from each subgroup in each cluster. (B) Heat map indicating top 5 marker genes expression for each cell cluster.

Figure 1

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